Use of anion exchange chromatography in weak partitioning mode to provide high empty AAV capsid removal and product yields

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Overview

- An anion exchange step to maximise full capsid enrichment for AAV-AQP1 (AAV2) was developed.
- The similar properties of the full and empty capsids led to a trade-off between product yield and full capsid ratio which made selecting optimal conditions difficult.
- Exploring the design space led to weak partitioning mode being established as a better alternative to bind-and-elute mode, and conditions that achieved the removal of >50% of the empty capsids were identified.
- As a result of the empty capsid removal, an eluate with a full capsid ratio of >80% (as confirmed by AUC) and a product yield >50% can be achieved.
- Understanding the design space led to the development of a robust and scalable process.

Key Results

Identifying Area of Interest

Understanding how the parameters interact meant an optimal region could be identified, allowing the key process objectives to be achieved.

Alternative Mode of Operation

Exploiting weak partitioning as an alternative to the typical bind-and-elute meant that almost **50%** of the empty capsids could be removed in the flow through alone. Coupled with a high NaCl wash, an eluate with >80% full capsids could be achieved.

1. Weak partitioning and where it is used

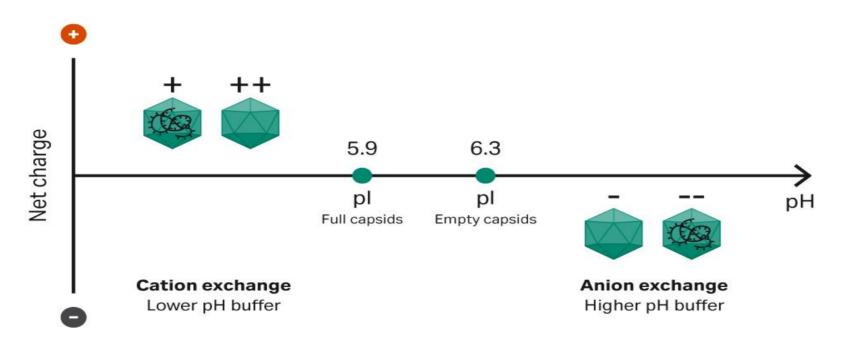


Figure 1. The charge of empty and full AAV capsids above and below the pl (1)

Weak partitioning describes the process of separating full capsids from empty capsids by creating an environment in which the binding of full capsids is more favourable.

- Anion exchange chromatography is a technique used to separate the full capsids from the empty capsids.
- This form of chromatography exploits the charge difference between capsids containing viral genome and those that are empty.
- Targeting pH values above the capsid pI means the VG containing full capsids carry a greater negative charge than the empty capsids.

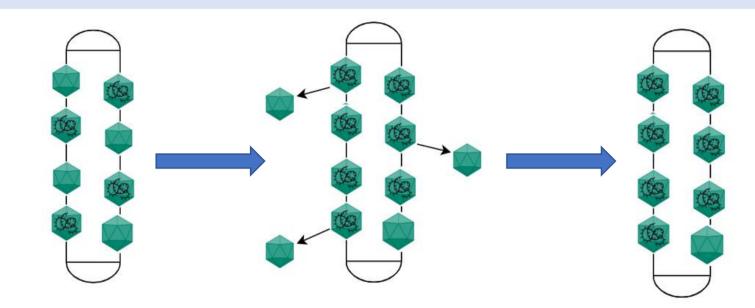


Figure 2: The weak partitioning process, where the empty capsids are replaced by full capsids due to the environment created

2. Initial design of experiments

Parameter

Load challenge (VG/mL)

Feed full capsid ratio (%)

Eq./feed NaCl conc. (mM)

Wash 2 NaCl conc. (mM)

Parameters and ranges for further optimization were identified from initial AEX platform-fit experiments.

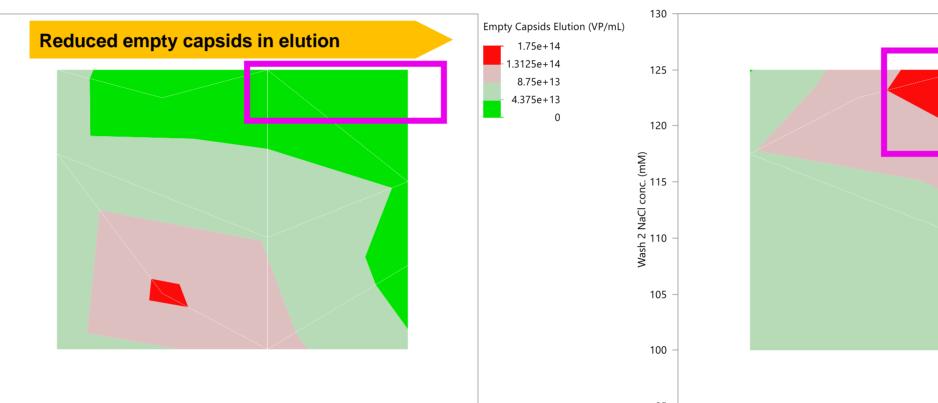
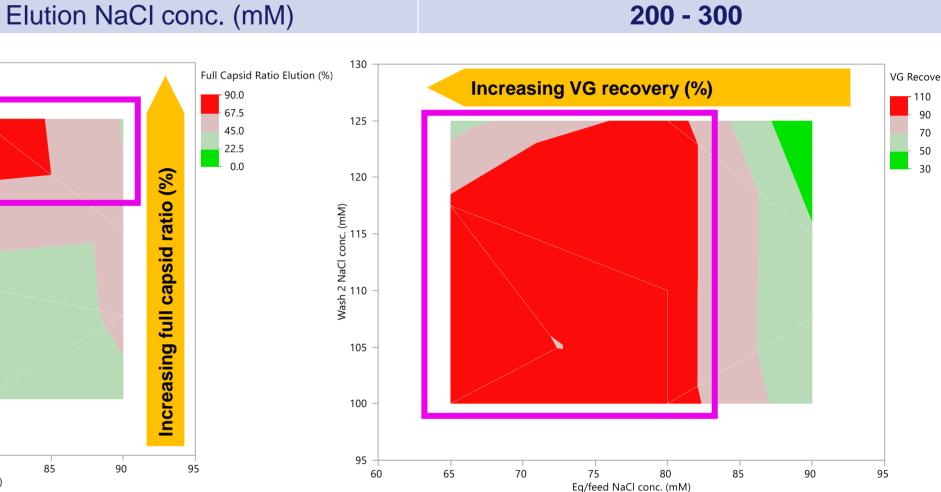


Figure 3: Relationship between wash 2 and eq. / feed NaCl concentrations (mM), and the empty capsids present in the elution (VP/mL)

Figure 4: Relationship between wash 2 and eq. / feed NaCl concentrations (mM), and the full capsid ratio (%) in the elution



Range

 $5 \times 10^{13} - 1 \times 10^{14}$

<10

65 - 90

100 - 125

Figure 5: Relationship between wash 2 and eq. / feed NaCl concentrations (mM), and the VG recovery (%) in the elution

Results of DOE combined to highlight optimal region within the design space, in line with the desired outcome.

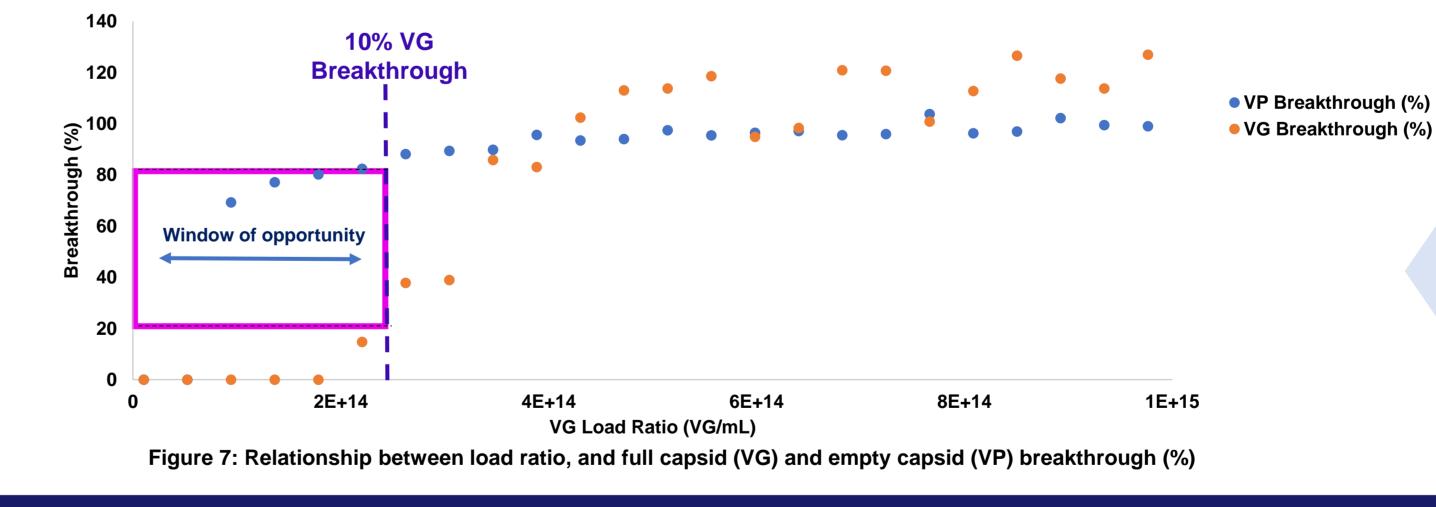
Figure 6: Optimal region based on the wash 2 and eq / feed NaCl concentrations (mM)

Value **Parameter** Empty capsid in **Greater than 50%** flowthrough **Empty capsid** Approx. 10% removal in wash 2 VG recovery in Approx. 50% elution Full capsid ratio in Approx. 50% the elution

3. Understanding how load ratio affects weak partitioning

The effects of weak partitioning are maximised by increasing the load ratio.

Parameter	Value		
Load ratio (VG/mL)	1 x 10 ¹⁵		
Eq. / feed NaCl conc. (mM)	90		



Exploiting the window of opportunity means up to 80% of empty capsids breaking through while only 10% of full capsids are lost in the flow through.

Value

4. Collating findings

Optimised Parameter Ranges Load ratio 1 x 10¹⁴ (VG/mL) Feed full capsid 31 ratio (%) Eq/feed NaCl 90 conc. (mM) Wash 2 NaCl 125 conc. (mM) Elution NaCl conc. 250 (mM)

Figure 4 indicated that for the largest empty capsid removal, high NaCl concentration was required for both conditions.

Increasing the Wash NaCl causes VG losses of up to 30%, 50% VG recovery is still recovered in the elution phase.

The process is scalable from

2L to 80L for both the VG

ratio.

recovery and the full capsid

Elution Wash 2 250.00 200.00 150.00 100.00 50.00 920.00 Volume (mL)

Figure 8: A chromatogram of an optimised AAV-AQP1 AEX step at 2 L scale

Results

by inversion

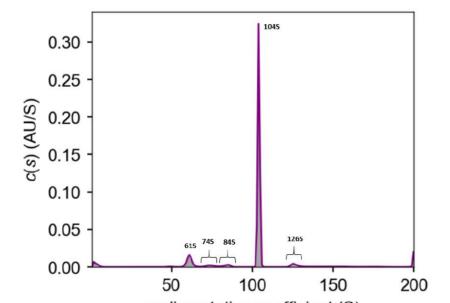
80% full capsid

ratio highlighted

Empty capsids in >40% flowthrough (%) Full capsids in <1% flowthrough (%) Full capsids in wash 33% 2 (%) Empty capsids in 20% wash 2 (%) VG recovery in **53%** elution (%) Full capsid ratio in 81.5% elution (%)

Parameter

5. Confirmation by AUC



sedimentation coefficient (S) Figure 9: An AUC result confirming the elution peak in Figure 8 contains 81.5% full capsids

6. Proven scalability

■ VG Recovery (%) Full Cansid Ratio (%)

					= Full Capsid Ratio (%)			
_	70							
8	60							
age	50							
ent	40							
Percentage	30							
_	20							
	10							
	0							
2 L Scale				Scale	80 L Scale			

Figure 10: A comparison between the full capsid ratio and VG recovery of a 2 L and 80 L batch

Conclusion

Understanding how to exploit weak partitioning is vital to maximise the full capsid enrichment. When coupled with an understanding of the process 'sweet spot' a process with a full capsid ratio of greater than 80% is achievable for AAV-AQP1.

References

1. Cytiva. 2022. Enhanced AAV downstream processing - Cytiva. [online] Available at: https://www.cytivalifesciences.com/en/us/solutions/cell-therapy/knowledge-center/resources/enhancedaav-downstream-processing.